EFFECT OF PENETRATION ENHANCERS ON TRANSDERMAL ABSORPTION OF INSULIN ACROSS HUMAN CADAVER SKIN V.U. Rao A.N. Misra* Pharmacy Department, Faculty of Technology & Engineering, M.S. University, Kalabhavan, Baroda, India.

ABSTRACT

transdermal diffusion οf insulin, polypeptide drug, across the human cadaver skin evaluated in vitro, in presence οf penetration enhancing solvents, anionic surfactants, biosurfactants, a natural moisturizing agent and combinations thereof. Also, attempt made relate the an was to penetration t.o physical parameters like distribution coefficient, surface tension and viscosity. The results permeation experiments indicate permeation enhancers used in the present investigation significantly enhance the amount of drug entering HCS and the amount reaching to the skin. permeation enhancement synergistic effect onwas permeation where observed in cases combination οf selectively used. Reasons enhancers were synergism were critically examined and established.

INTRODUCTION

several There have been attempts at delivering therapeutic peptide and protein drugs by nonparenteral routes including oral, buccal, rectal and transdermal Recent reports revealed transdermal permeation (1-4).macromolecules using penetration enhancers dimethyl sulfoxide, azone and by physical means such as iontophoresis, streaming hot air flow, etc. Considering the fact that the transdermal drug delivery possibility οf bypassing the provides the gastrointestical first pass elimination and of achieving a better patient compliance, it is therefore logical to consider it as a potential route for delivery of peptide based pharmaceuticals.



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The aim of the present study was to evaluate in effect of penetration enhancing the anionic biosurfactants surfactants, and a natural moisturizing agent on the transdermal diffusion insulin, model polypeptide drug across HCS. establish whether physical parameters are responsible enhancement i.n penetration, distribution coefficient, surface tension and viscosity determined.

EXPERIMENTAL

24.8 IU/mg preparation οf insulin (Novo, Denmark) was used. Dimethyl sulfoxide (DMSO), dymethylformamide (DMF), sodium deoxycholate sodiumtaurocholate, sodiumtauroglyco-cholate (NTGC), sodiumphosphate, potasium di hydrogen orthophosphate and sodium chloride (National chemicals, India), Pluronic F68 (PF68), Pluronic F127 (PF127) (BASF, USA) were used.

Reagents:

- pH 7.4 phosphate buffered saline (PBS): This buffer was prepared as per B.P. 1980.
- Stock solution of insulin: prepared by dissolving 50mg of drug PBS.
- Stock solutions οf the surfactants: These prepared by dissolving 50mg of the powder in 10ml
- Formulations: Each formulation contained 800ug/ml of insulin and specified concentration of penetration enhancer/s as shown in Table-1

Methods:

In vitro permeation tests:

permeation study was conducted by applying 250µl of each formulation on the epidermal surface of a piece of HCS tied at one end of a glass permeation tube. The tube was dipped flush to the surface of 25ml contained in a receptor compartment maintained at 37.5 ± 0.5°C and strirred at 50rpm with a magnetic stirrer. 2ml aliquots were withdrawn at each sampling interval from the receptor cell and insulin spectrophtometrically at UV-visible 2000 276mm on Hitachi Spectrophtometer. positive Negative and blanks were also simultaneously to ensure noninterference leachings and the permeation enhancers. 2ml fresh PBS was added after each withdrawal. A 1 1 permeation runs and sample analysis were carried out in triplicate over a period of 6 hours.



TABLE-1

(+ = present = absent.) Formu- 30% 30% 0.01% 0.01% 0.1% 0.1% 0.1% 0.1% 10% 10% 1ation DMF DMSO PF127 PF68 NDC NTC NTGC Urea No. Group A A1	Formulations Containing Penetration Enhancers			Insulin		(40 IU/ml))		and	
Iation DMF DMSO PF127 PF68 NDC NTC NTGC Urea Group A A1 + -			(+=	present		= abs	ent.)		
Iation DMF DMSO PF127 PF68 NDC NTC NTGC Urea Group A A1 + -	_								
Group A A1 + - <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>									
A1 + - <t< td=""><td></td><td>DMF</td><td>DMSO</td><td>PF12/</td><td>P1.08</td><td>NDC</td><td>NTC</td><td>NTGC</td><td>Urea</td></t<>		DMF	DMSO	PF12/	P1.08	NDC	NTC	NTGC	Urea
A1 + - <t< td=""><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>									
A2 - + A3 +									
A3 +		+	-	-	-	-	-	-	-
A4		-	+	-	-	-	-	-	-
A5 + A6 + A7 + A8 + + + +		_	-	+	_	-	-	-	-
A6 + + A8 + + + + + + + +		-	_	-	+	-	-	-	-
A7		-	-	_	-	+	-	-	
A8 + Group B B1 - + - +		-	-	_	_	_	+	-	-
Group B B1 - + - + B2 - + + B3 - + + - Group C C1 + - + - + C2 + + - +		-	-	_	_	-	-	+	-
B1 - + - + B2 - +	A 8	-	-	-	-	-	_	-	+
B2 - + + + + + + + + + + + + + +	Group B								
B3 - + + - Group C C1 + - + - + - + C2 - + - + + - +		-	+	_	+	_	-	-	-
Group C	B2	-	+	-	_	-	+	-	-
C1 + - + + C2 - + - + - + - +	В3	-	+	-	-	-	-	+	-
C1 + - + + C2 - + - + - + - +	Group C								
C2 + - +	CI	_	_	_	+	-	-	-	+
C3 + +		_	-	-	_	_	+	-	+
	C3	_ 	-		- 	- 	-	+	+

Determination οf apparent distribution viscosity and surface tension:

the determination of apparent distribution For coefficient, the HCS was cut in circular discs of 2 cm and put in glass vials each containing 10ml of solution. Each solution contained lmg of insulin with composition of penetration enhancers as shown in Table-The vials were shaken thoroughly at 37 ± 0.5 °C for 4 hours, centrifuged at 3000 rpm for 20mins after 24 hours and the supernatants were analysed.

Surface tension and single point viscosity of each formulation was determined using Stalagmometer and Ostwald Viscometer respectively. Negative and positive blanks were run for all measurements. Each measurement was carried out in triplicate.



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DATA ANALYSES

The experimental model adopted here is a simple flux case. The quantity of drug which order permeates the HCS, Q'(t) and the quantity which passes through the HCS and reaches the sink Q°(t) are the factors varying with time. These were calculated using equations previously reported (6). The rate at which the macromolecule enters into the skin $(dQ^{\circ}(t)/dt)$ and the rate which it enters into the sink $(dQ^{\circ}(t)/dt)$ were calculated from the slopes of the regressed lines of $Q^{1}_{(+)}$ vst and $Q^{\circ}_{(+)}$ vst values. The data of the of $Q^1_{(t)}$ vst and $Q^{\circ}_{(t)}$ vst values. The data of the apparent distribution coefficient, surface tension, viscosity, $dQ^1_{(t)}/dt$ and $dQ^{\circ}_{(t)}/dt$ values for each formulation are given in Table-2.

Statistical analysis using ANOVA was applied to the data of $Q^1(t)$, $Q^0(t)$, $dQ^1(t)$ /dt and $d^0(t)$ /dt values. The data was considered significant for p 0.05.

RESULTS AND DISCUSSION

In case of 30% DMSO there is a significant enhancement in the $Q^1(t)$ and $Q^\circ(t)$ values after 60 minutes with a 2 fold enhancement in the $dQ^{1}(t)/dt$ value and 2.3 fold enhancement in $dQ^{\circ}(t)/dt$ value as compared to the control formulation. In case of DMF, statistically significant enhancement occures after 90 minutes with 2 fold enhancement in dQ°(t)/dt value and 2.22 fold enhancement in the dQ°(t)/dt value. DMF significantly 30% DMSO and 30% enhance viscosity of the formulations which may be contributory to the increased osmotic activity leading to distortion of the laminated structure of the stratum corneum (8). The effect of DMF is statistically less significant as compared to DMSO which may be because the transdermal water loss caused by DMF is reported to be less intense and of shorter duration as compared to DMSO (9).

Formualtions containig nonionic surfactants PF27 and PF68 show a significant enhancement in the $Q^{1}(t)$ and $Q^{\circ}(t)$ values at all sampling time intervals. The significantly higher $dQ^{1}(t)/dt$ and $dQ^{\circ}(t)/dt$ values for PF68 as compared to PF127 indicate that PF68 better penetration enhancer for insulin as compared to PF127, One reason may be that PF68 brings significantly more reduction in the surface tension of the system as compared to the PF127 (Table-2).

All the three biosurfactants studied demonstrated a significant increase in the $Q^1(t)$ and $Q^{\circ}(t)$ values at all time intervals as compared to the control formulation. NTC and NTGC show a 2.27 and 2,42 folds enhancement in $dQ^1(t)/dt$ value respectively and 2.43 and 2.8 times enhancement in $dQ^{\circ}(t)/dt$ value. On



TABLE-2

The Data for $dQ^{i}_{(t)}/dt$, $dQ^{o}_{(t)}/dt$, Apparent Distribution Coefficient, Viscosity and Surface Tension for the Formulations containing Penetration Enhancers

Formu- lation No.	dQ ⁱ (t) dt x 10 ⁻³	dQ° (t) dt x 10 ⁻³	Apparent Distri- bution Coeffi- cient	Visco- sity (Cps)	Surface Tension dynes/ cm
0	4.01	3.50	0.073	0.787	68.360
A1 A2 A3 A4 A5 A6 A7 A8 B1 B2 B3 C1 C2	8.04 8.50 7.10 8.80 8.80 9.10 9.80 8.30 25.50 39.90 42.70 18.60 28.00 31.50	7.80 7.10 7.20 8.20 8.40 8.50 9.70 7.50 20.70 36.50 38.70 17.20 26.00 30.60	0.082 0.086 0.104 0.122 0.129 0.132 0.134 0.096 0.162 0.194 0.196 0.164 0.162 0.168	1.134 1.255 0.902 0.868 0.879 0.837 0.845 0.845 1.361 1.673 1.540 0.873 0.950	66.837 65.081 51.907 44.024 36.994 34.480 29.099 67.168 43.022 32.172 28.325 51.654 36.215 30.567

B2 = 30% DMSO + 0.1% NTGC

C2 = 10% Urea + 0.1% NTGC

C3 = 10% Urea + 0.1% PF68

B3 = 30% DMSO + 0.01% PF68

Key 0 = Control Formulation Bl = 30% DMSO + 0.1% NTC A1 = 30% DMFA2 = 30% DMSO B3 = 30% DMSO + 0.01% PF C1 = 10% Urea + 0.1% NTC A3 = 0.01% PF127A4 = 0.01% PF68

A5 = 0.1% NDCA6 = 0.1% NTC

A7 = 0.1% NTGCA8 = 10%Urea

the other hand, NDC shows only a 2.20 fold increase in $dQ^{+}(t)/dt$ and 2.34 fold increase in $dQ^{\circ}(t)/dt$ values. Among the three biosurfactants, NDC reduces the surface This coupled with the fact that tension least. number increases and hence aggregation solubility decrease at pH 7.4 must be responsible for the least activity.



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Urea, a natural moisturing agent show significant $Q^{1}(t)$ and Q°(t) in the values after minutes as compared to the control formulation. There is a 2.07 fold values. From Table-2 it is apparent that permeation enhancing property of urea cannot attributed to its ability to modify the surface tension and viscosity of the formulation but may be due to its moisturising and keratolytic activity.

The penetration enhancing solvents, surfactants show a significant enhancement permeation of insulin. Since each of these agents act by different mechanisms, it was deemed worthwhile study the penetration enhancing activity formulations containing combinations οf surfactants and urea and surfactants (Table-1). As compared to the control formulation enhancement $dQ^{-}(t)/dt$ value is 6.37 fold for formulation B1, 9.97 fold for formulation B2 and 10.67 times for formulation B3. The $dQ^{1}(t)/dt$ value increase 2.89, 2.80 and 2.13 folds for formualtions Bl, B2 and B3 respectively as compared to formualtions A4, A6 and A7.

In case of formulation Cl, C2 and C3 there is a 3.80 fold increase in the $dQ^{1}(t)/dt$ value 3.08 respectively as compared to formualtions A4, A6 and A7.

Similar enhancement is observed in case dQ°(t)/dt indicating values that both, the rate which the drug penetrates the skin and rate at which drug enters into the sink are enhanced in case of the combination.

in the case οf combination οf DMSO viscosity srufactants the οf the formualtions significantly enhanced and the surface tension is also reduced to an extent which is greater than that formualtions conataining single agents. Thus increased osmotic activity coupled with the reduced thermodynamic activity may be the reason synergistic effect. The ability of urea to hold water coupled with the reduction in the interfacial brought about by the surfactants may be the reason for the synergstic activity in case of formualtions Cl to C3.

CONCLUSIONS

All the penetration enhancers used in the present investigation significantly enhance the amount of drug entering into the HCS and the amount of drug reaching of permeation sink. The extent enhancement from one enhancer to other and was found to follow the rank order:

B3 > B2 > C3 > C2 > B1 > C1 > A7 > A6 > A5 >A4 > A8 > A2 > A3 > A1 > 0



Physical parameters such as distribution coefficient, surface tension and viscosity were for explaining the mechanism of permeation enhancement by different classes of enhancers and also can helpful in predicating the degree of enhancement that class. A synergistic effect was observed in cases where combination of enhancers were selectively is assumed that to acheive degree same enhancement the toxicities will be comapratively less in case of use of selective combinations as compared to penetration enhancers. However, the combinations οf enhancers can only be settled after biological investigation first on animal subjects. Further efforts followed by human concentrate on use οf selective combinations penetration enhancers in the development of transdermal delivery system for polyeptide drugs end evaluation of their toxicities.

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